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The inflammation, vascular repair and injury responses to exercise in fit males with and without Type 1 diabetes: an observational study

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Abstract

Background: Type 1 diabetes is associated with raised inflammation, impaired endothelial progenitor cell mobilisation and increased markers of vascular injury. Both acute and chronic exercise is known to influence these markers in non-diabetic controls, but limited data exists in Type 1 diabetes. We assessed inflammation, vascular repair and injury at rest and after exercise in physically-fit males with and without Type 1 diabetes.

Methods: Ten well-controlled type 1 diabetes (27 ± 2 years; BMI 24 ± 0.7 kg.m²; HbA_{1c} 53.3 ± 2.4 mmol/mol) and nine non-diabetic control males (27 ± 1 years; BMI 23 ± 0.8 kg.m²) matched for age, BMI and fitness completed 45-min of running. Venous blood samples were collected 60-min before and 60-min after exercise, and again on the following morning. Blood samples were processed for TNF- α using ELISA, and circulating endothelial progenitor cells (cEPCs; CD45^{dim}CD34⁺VEGFR2⁺) and endothelial cells (cECs; CD45^{dim}CD133⁻CD34⁺CD144⁺) counts using flow-cytometry.

Results: TNF- α concentrations were 4-fold higher at all-time points in Type 1 diabetes, when compared with control ($P < 0.001$). Resting cEPCs were similar between groups; after exercise there was a significant increase in controls ($P = 0.016$), but not in Type 1 diabetes ($P = 0.202$). cEPCs peaked the morning after exercise, with a greater change in controls vs. Type 1 diabetes (+139 % vs. 27 %; $P = 0.01$). cECs did not change with exercise and were similar between groups at all points ($P > 0.05$). Within the Type 1 diabetes group, the delta change in cEPCS from rest to the following morning was related to HbA_{1c} ($r = -0.65$, $P = 0.021$) and TNF- α ($r = -0.766$, $P = 0.005$).

Conclusions: Resting cEPCs and cECs in Type 1 diabetes patients with excellent HbA_{1c} and high physical-fitness are comparable to healthy controls, despite eliciting 4-fold greater TNF- α . Furthermore, Type 1 diabetes patients appear to have a blunted post-exercise cEPCs response (vascular repair), whilst a biomarker of vascular injury (cECs) remained comparable to healthy controls.

Keywords: TNF- α , Endothelial progenitor cells, Circulating endothelial cells

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Background

Exercise carries many health benefits for people with Type 1 diabetes [1]. Of particular importance, regular physical exercise is associated with improvements in an array of cardiovascular risk factors such as cardiorespiratory fitness, endothelial function, and blood lipid profiles. These benefits are of significance given the raised cardiovascular risk and early mortality in these patients [2].

Exposure to hyperglycaemia and chronic inflammation ultimately means that individuals with Type 1 diabetes demonstrate raised markers of vascular damage, such as circulating endothelial cells (cECs; [3]), and endothelial dysfunction [4], in comparison with healthy controls [4, 3]. Accompanying this may be a reduction in circulating endothelial progenitor cell count (cEPCs; [4, 5]); bone marrow derived cEPCs are highly important for vascular repair and protection [5, 6] and are a significant predictor of endothelial function [4, 7] and cardiovascular risk [7].

A single exercise bout has been shown to acutely raise cEPCs count in both healthy and patient populations [8–11]. For example, Rehman et al [8] demonstrated that a single bout of cycling exercise significantly raised cEPCs by ~4 fold after exercise, in sedentary overweight males. Similar increases in cEPCs after exercise have been demonstrated in healthy, physically fit individuals, as well as in sedentary individuals with chronic disease [9]. However, data on the acute cEPCs response to exercise within Type 1 diabetes is lacking. Regular exercise training is associated with raised resting cEPCs and has a strong anti-inflammatory effect [12, 13]. Indeed, these benefits are likely integral to the cardio-protective role regular exercise provides. With this in mind, and the association of Type 1 diabetes with reduced cEPCs and raised inflammation, it is of scientific importance to determine the acute cEPCs response to exercise within Type 1 diabetes patients.

Moreover, how the resting cEPCs, cECs and inflammatory status of physically fit type 1 diabetes patients compares to matched, non-diabetic controls would also be of interest. Physical fitness is a measure that is not considered within the existing literature comparing cEPCs (marker of vascular repair), cECs (marker of vascular damage) and inflammation in this population [4, 3]. Therefore, the aim of this study was to assess the cEPCs, cECs and inflammation at rest and in response to submaximal exercise in physically-fit males with and without Type 1 diabetes.

Methods

Participants

Eligibility criteria for type 1 diabetes patients consisted of being aged between 18 and 35 years, a duration of diabetes > 2 years, and an HbA_{1c} < 8.0 % (64 mmol/mol). In addition, patients were required to be free of all diabetes-related complications including impaired awareness of hypoglycaemia

Table 1 Type 1 diabetes and control group participant characteristics mean ± SEM

	T1DM	CON	P value
Age (years)	27 ± 2	27 ± 1	=0.798
BMI (kg.m ²)	24 ± 0.7	23 ± 0.8	=0.243
VO ₂ max (ml.kg.min ⁻¹)	51.0 ± 2.1	50.7 ± 1.1	=0.808
HbA _{1c} (mmol/mol / %)	53.3 ± 2.4 / 6.9 ± 0.2 %	—	—
Diabetes duration (years)	12 ± 2.0	—	—

P calculated from independent samples t-test
BMI Body Mass Index, VO_{2max} aerobic capacity

(assessed via the Clarke method [14]) and not receiving any medication other than insulin. Participants in both the Type 1 diabetes and control group had to be regularly and consistently undertaking exercise (participating in aerobic based exercise for a minimum of 30 min at a time, at least three times per week, for >12 months). Smokers were excluded from both groups. Ten males with type 1 diabetes and nine non-diabetic male controls, matched for age, anthropometry, and fitness were recruited for this study (Table 1). Patients were treated with a basal-bolus regimen composed of long-acting insulins glargine (*n* = 8) or detemir (*n* = 2), and rapid-acting insulin aspart. All patients were stable on their respective insulin regimen for a minimum of 1 year, and were familiar with carbohydrate counting, administering 1.0 ± 0.2 units of insulin aspart (IU) per 10 g of carbohydrate.

Preliminary testing

Fully informed written consent was obtained from all participants following the study's approval from local National Health Service Research Ethics Committee (13/NE/0016). Type 1 diabetes participants attended the Newcastle National Institute for Health Research Clinical Research Facility exercise laboratory for a preliminary screening visit as described previously in detail [15], before returning to establish peak cardio-respiratory parameters during the completion of an incremental-maximal treadmill running protocol as per the methods of Campbell et al [15], to determine the individual speed participants would run at during the experimental trial. The control group completed the same preliminary tests at the Exercise Physiology laboratory at Northumbria University.

Experimental design

Type 1 diabetes patients were fitted with a real-time continuous glucose monitor (Paradigm Veo, Medtronic diabetes, Northridge, CA, USA) >24 h prior to the laboratory visit with high and low alerts set to help maintain glycaemia within normal ranges prior to the experimental session. Participants did not exercise within 96 h of the experimental visit.

On the experimental day participants were provided with standardised cereal-based breakfast and pasta-based lunch; prescribed by the research team. Participants arrived to the laboratory at 17:00 h. Following a resting blood sample participants consumed a pre-exercise carbohydrate bolus (corn flakes, peaches, semi-skimmed milk; 1738 ± 71 kJ / 415 ± 17 kcal) equating to 1.0 g.carbohydrate.kg⁻¹ body mass. With this meal, patients self-administered a 25 % (2.0 ± 0.5 IU) dose (i.e. a 75 % reduction [15–19]) of rapid-acting insulin into the abdomen. Following this bolus, participants remained rested for 60 min, before commencing 45 min of treadmill running at a speed calculated to elicit 70 % of VO_{2max} . This exercise intensity falls within current recommendations of the American College of Sports Medicine [20]. All participants completed the exercise protocol and there were no hypoglycemic episodes within the Type 1 diabetes group.

After exercise, participants remained at rest for 60 min before a further blood sample was collected. Participants were then discharged from the laboratory. On the following morning, participants returned to the laboratory at 08:00 h for a further resting, fasted blood sample.

Blood sampling and data analysis

Venepuncture technique was used to collect 10 ml of whole blood at each respective sample point. After discarding the first four ml of collected blood, samples were evenly dispersed into a K2EDTA and lithium heparin tube. The lithium heparin tube was centrifuged for 15 min at 3000 rpm (4 °C) and stored at -80 °C for retrospective analysis of plasma Human TNF- α (Quantikine ELISA, R&D Systems, Roche Diagnostics, West Sussex, UK). The K2EDTA was sent for analysis of cEPCs and cECs immediately and was processed within 2 h [21].

Circulating endothelial progenitor cells and circulating endothelial cells

Flow cytometry

100 μ l of whole blood was incubated with 5 μ l of V500 CD45 (BD Biosciences, Oxford, UK), 20 μ l of PerCP-Cy5.5 CD34 (BD Biosciences, Oxford, UK), 5 μ l of PE VEGFR-2+ (R&D Systems, Roche Diagnostics, West Sussex, UK), 5 μ l APC CD133 (BD Biosciences, Oxford, UK), 10 μ l FITC CD144 (BD Biosciences, Oxford, UK) for 30 min. Subsequently, 2 ml of pharmlyse (BD Biosciences, Oxford, UK) was used to lyse the red cells. The sample was then analysed by flow cytometry on BD FACS Canto™ II system and samples were run to approximately 500,000 total events. Analysis was carried out using BD FACSDiva™ software. On average 440,000 events were counted. CEPCs events were defined using the most recent definition of CD45^{dim}CD34⁺VEGFR2 (KDR)⁺, as recommended by Van Craenenbroeck et al. [21]. Intra-assay variation (CD45^{dim}CD34⁺VEGFR-2⁺)

was less than 8 %. The results were expressed as % leukocytes [22, 21]. CECs events were defined as CD45^{dim}CD133⁻CD34⁺CD144⁺ [23, 24].

Statistical analysis

Statistical analysis was performed using PASW Statistics 18 software (IBM, Armonk, NY) with significance set at $p \leq 0.05$. Plasma TNF- α responses were analysed using mixed model ANOVA (group*time) and are presented as mean \pm SEM. CEPCs and CECs responses were assessed using two-way Friedman's analysis and are presented as median \pm IQR. Between group differences, and comparisons of the delta-change in cEPCs and cECs counts, were assessed using Mann-Whitney U test. Relationships were assessed with Pearson's product moment correlation coefficient or Spearman's rank order correlation coefficient.

Results

TNF- α

Plasma TNF- α responses are presented in Fig. 1. There was no group*time interaction ($P = 0.324$), or change over time ($P = 0.103$), however there was a significant effect of group ($P < 0.001$). The Type 1 diabetes group elicited ~4 fold greater TNF- α concentrations at all sample points, when compared to control (Fig. 1).

CEPCs & cECs

CEPCs counts are presented in Fig. 2a. Baseline cEPCs counts were similar between groups (Fig. 2a). There was a significant increase in cEPCs counts after exercise within the control group ($P = 0.016$), however, there was no change within the Type 1 diabetes patients ($P = 0.202$). CEPCs counts peaked the morning after exercise within the control group (Fig. 2a) and the delta change, from

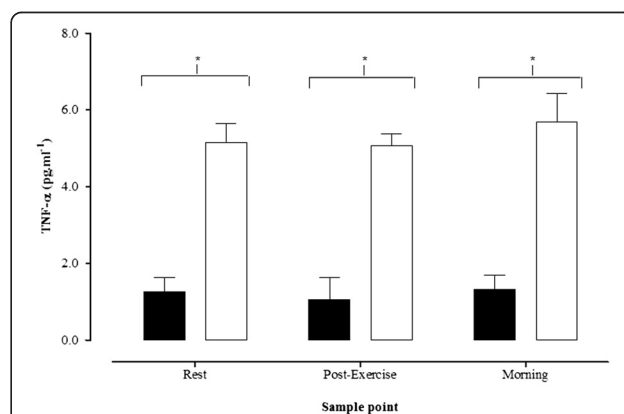


Fig. 1 Plasma TNF- α at rest, 60 min post-exercise, and on the morning after exercise. Solid black bars = Control group, solid white bars = Type 1 diabetes group. * indicates a significant between group difference ($P < 0.05$). Data presented as mean \pm SEM

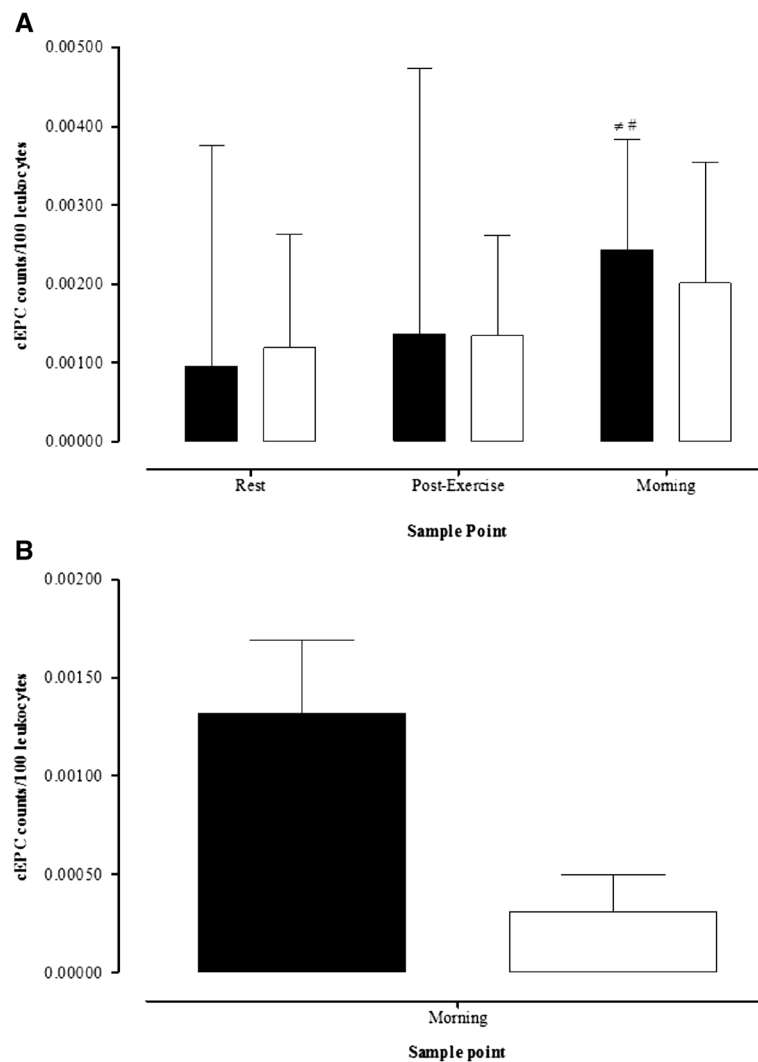


Fig. 2 a Circulating endothelial progenitor cell (cEPC) counts at rest, 60 min post-exercise, and on the morning after exercise. Solid black bars = Control group, solid white bars = Type 1 diabetes group. # indicates within conditions difference to pre-exercise, # indicates within condition difference to post-exercise ($P < 0.05$). **b** Change in cEPCs counts from rest to the morning after exercise. Solid black bars = Control group, solid white bars = Type 1 diabetes group. Data presented as median \pm IQR

baseline to morning, tended to be greater within the control group (Fig. 2b; $P = 0.09$). The median cEPCs count increased by 139 % and 27 % in the control and type 1 diabetes groups, respectively ($P = 0.01$). CECs count responses are presented in Fig. 3. CECs count was similar at all-time points between groups, and there was no significant change in cECs with exercise within either Type 1 diabetes ($P = 0.179$) or control ($P = 0.236$) groups.

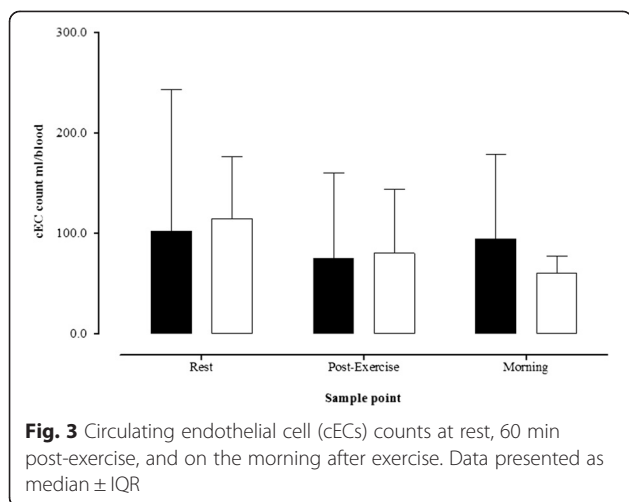
Relationships between variables

Within the Type 1 diabetes group, the delta change in cEPCs from rest to the following morning was related to HbA_{1c} ($r = -0.65$, $P = 0.021$) and TNF- α ($r = -0.766$, $P = 0.005$). There were no other significant relationships

between cEPCs, cECs and TNF- α and participant characteristics (VO_{2peak} , BMI, age and duration of diabetes) in either the control or Type 1 diabetes group.

Discussion

This study aimed to examine cEPCs, cECs and TNF- α at rest, and in response to acute submaximal exercise, in physically fit males with and without Type 1 diabetes. This study is the first of this kind and we have shown that resting cEPCs and cECs were similar between groups, despite patients demonstrating ~ 4 fold greater TNF- α concentrations. However, the increase in cEPCs was blunted in the patients, with the change from rest to the morning following exercise being ~ 112 % lower in



Type 1 diabetes, when compared to control participants. Furthermore, our data showed that this blunted response within the Type 1 diabetes group appeared to be predicted by HbA_{1c} and TNF- α concentrations.

Resting cEPCs and cECs

Our finding that resting cEPCs and cECs were similar between groups is in agreement with Fadini et al. [25], but also contrasts with other literature in the area [4, 3]. In comparison with Sibal et al. [4], the Type 1 diabetes patients of this study were of comparable age and had similar duration of diabetes and also shared a raised inflammatory state. The difference in the findings are potentially explained by the high cardio-respiratory fitness and excellent glycemic control of our patients (HbA_{1c} \sim 7 %, 53 mmol/mol vs. \sim 8.5 %, 69.4 mmol/mol [4]). Indeed, the participants in the study of Fadini et al. [25] also had good HbA_{1c} (\sim 7.7 %). Whilst the relationship between glycemic control and cEPCs is established in Type 2 diabetes and acute myocardial infarction [26, 27], there are few studies in Type 1 diabetes. Most recently it has been shown that in children with Type 1 diabetes, cEPCs count was inversely related with HbA_{1c} [28]. In our study, it should be reassuring to both patients and clinicians that after achieving good glycemic control cEPCs were comparable to healthy controls. However, since our patients were also very fit, it would be interesting to investigate the change in HbA_{1c} and cardio-respiratory fitness both independently, and concomitantly (e.g. via an exercise training programme), to provide further insight into the normalised cEPCs we show.

cEPCs response to exercise

This is the first study to examine acute exercise and cEPCs regulation in Type 1 diabetes and our findings suggest that regulation of cEPCs release in response to an exercise stimulus may be abnormal despite eliciting normal resting cEPCs and excellent glycemic control. Research

examining the bone marrow biology of both animals and patients with diabetes confirms that bone marrow function is impaired [29] which may reduce cEPCs mobilisation [25]. Recent research has shown that the mobilisation of cEPCs in response to administration of recombinant granulocyte colony-stimulating factor is impaired in diabetes patients [25]. Speculatively, bone marrow function may also have contributed to the blunted cEPCs response to exercise demonstrated in the Type 1 diabetes patients.

It is well established that resting cEPCs are normal or high in active individuals [13, 9], thus it is of interest that both groups of participants in our current study had a cardio-respiratory fitness (VO_{2max} 50 ml.kg.min⁻¹) which would categorise them as excellent, or above average (average VO_{2max} for males aged \sim 27 being \sim 42 ml.kg.min⁻¹ [20]). Earlier research has described \sim 4 fold higher cEPCs in trained marathon runners when compared to sedentary control participants [13], and there is evidence that regular aerobic-exercise training is an effective intervention to raise cEPCs in both healthy [13] and patient populations [9, 30]. Potentially, the regular aerobic exercise engaged in by the participants in this study, evidenced by high cardio-respiratory fitness, may additionally explain the comparable resting cEPCs shown.

Inflammation and cEPCs

Our study provides another interesting finding that the comparable cEPCs and cECs between groups were concomitant with \sim 4 fold higher TNF- α concentrations seen in our Type 1 diabetes patients. The raised inflammation is in agreement with our previous data of Sibal et al. [4]. There is evidence, in advanced cardiovascular disease patients, that cEPCs are reduced potentially due to the myelosuppressive effects of TNF- α [31]. Furthermore, three months of TNF- α inhibitory drug treatment has been shown to significantly raise cEPCs in rheumatoid arthritis patients [32]. Further research is required to investigate the interaction between the anti-inflammatory effects of regular exercise in populations which experience chronic inflammation, such as Type 1 diabetes.

We show for the first time that despite the excellent glycemic control and high physical fitness of the Type 1 diabetes patients there was a blunting of the rise in cEPCs elicited after exercise. Shear stress, increased nitric oxide (NO) production, through increased activity of endothelial nitric oxide synthase, and hypoxia are suggested to be key stimuli that contribute to the mobilisation of cEPCs with exercise [33, 9]. However, in some instances oxidative stress can reduce NO availability, which could, speculatively, attenuate the signal for cEPCs mobilisation. Recent research has shown that well controlled Type 1 diabetes patients demonstrate increased oxidative stress during aerobic exercise, in comparison with non-diabetic control participants [34].

Furthermore, there may also be a role for the myelosuppressive effect of TNF- α in these responses. Future research should examine the role of both oxidative stress and inflammatory signalling in the acute cEPCs response to exercise.

Although this study is limited by a relatively small sample size, it is important to consider the homogenous group that were observed, with patients all male, aged 19–34 years, HbA_{1c} range of 6–7.9 %, and all physically fit; as such, the importance of these data should not be underestimated. Future research should explore how improving HbA_{1c} independently of improving physical fitness influences markers of vascular repair at rest and in response to exercise. Conversely, it would be of interest to explore if improving physical fitness can rescue the deleterious effect of poor glycaemic control on cEPCs regulation. Furthermore, the role of the insulin species patients are treated with could be an additional factor to consider when examining the impact of exercise on cEPCs in this population [35].

Conclusions

In conclusion, Type 1 diabetes patients with high cardiorespiratory fitness and excellent glycaemic control present normal resting markers of vascular repair (cEPC's) and injury (cECs), despite being in a persistent inflammatory state (raised TNF- α). However, the cEPCs response to acute exercise appears blunted, potentially limiting the cardiovascular benefits of exercise.

Abbreviations

EPC: Response to exercise in Type 1 diabetes; cEPC: Circulating endothelial progenitor cell; cEC: Circulating endothelial cell; HbA_{1c}: Glycated haemoglobin; TNF- α : Tumour Necrosis Factor Alpha; VO_{2max}: Maximum oxygen uptake.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MDC, JUW, JG, FA, contributed to the conception and design of the study and researched data and reviewed/edited the manuscript. MW & JAS aided in participant recruitment, data analysis and reviewed/edited the manuscript. FWA, SW performed flow cytometry, data interpretation and reviewed/edited the manuscript. EJS aided in data collection and reviewed/edited the manuscript. DJW contributed to the conception, design, data collection and wrote the manuscript. DJW is the guarantor of this work and, as such, had full access to all of the data in the study and takes full responsibility for the integrity of the data and the accuracy of the data analysis. All authors read and approved the final manuscript.

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References

1. Chu L, Hamilton J, Riddell MC. Clinical management of the physically active patient with type 1 diabetes. *Phys Sportsmed*. 2011;39:64–77.
2. de Ferranti SD, de Boer IH, Fonseca V, Fox CS, Golden SH, Lavie CJ, et al. Type 1 diabetes mellitus and cardiovascular disease: a scientific statement from the American Heart Association and American Diabetes Association. *Diabetes Care*. 2014;37:2843–63.
3. Ascioglu E, Gogas Yavuz D, Koc M, Ozben B, Yazici D, Deyneli O, et al. Circulating endothelial cells are elevated in patients with type 1 diabetes mellitus. *Eur J Endocrinol*. 2010;162:711–7.
4. Sibal L, Aldibbiat A, Agarwal SC, Mitchell G, Oates C, Razvi S, et al. Circulating endothelial progenitor cells, endothelial function, carotid intima-media thickness and circulating markers of endothelial dysfunction in people with type 1 diabetes without macrovascular disease or microalbuminuria. *Diabetologia*. 2009;52:1464–73.
5. Fadini GP, Ferraro F, Quaini F, Asahara T, Madeddu P. Concise review: diabetes, the bone marrow niche, and impaired vascular regeneration. *Stem Cells Transl Med*. 2014;3:949–57.
6. Hernandez SL, Gong JH, Chen L, Wu IH, Sun JK, Keenan HA, et al. Characterization of circulating and endothelial progenitor cells in patients with extreme-duration type 1 diabetes. *Diabetes Care*. 2014;37:2193–201.
7. Hill JM, Zalos G, Halcox JP, Schenke WH, Waclawiw MA, Quyyumi AA, et al. Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. *N Engl J Med*. 2003;348:593–600.
8. Rehman J, Li J, Parvathaneni L, Karlsson G, Panchal VR, Temm CJ, et al. Exercise acutely increases circulating endothelial progenitor cells and monocyte-/macrophage-derived angiogenic cells. *J Am Coll Cardiol*. 2004;43:2314–8.
9. Volaklis KA, Tokmakidis SP, Halle M. Acute and chronic effects of exercise on circulating endothelial progenitor cells in healthy and diseased patients. *Clin Res Cardiol*. 2013;102:249–57.
10. Thorell D, Borjesson M, Larsson P, Ulfhammer E, Karlsson L, DuttaRoy S. Strenuous exercise increases late outgrowth endothelial cells in healthy subjects. *Eur J Appl Physiol*. 2009;107:481–8.
11. Van Craenenbroeck EM, Vrints CJ, Haine SE, Vermeulen K, Goovaerts I, Van Tendeloo VF, et al. A maximal exercise bout increases the number of circulating CD34+/KDR+ endothelial progenitor cells in healthy subjects. Relation with lipid profile. *J Appl Physiol*. 2008;104:1006–13.
12. Palmefors H, DuttaRoy S, Rundqvist B, Borjesson M. The effect of physical activity or exercise on key biomarkers in atherosclerosis—a systematic review. *Atherosclerosis*. 2014;235:150–61.
13. Bonsignore MR, Morici G, Santoro A, Pagano M, Cascio L, Bonanno A, et al. Circulating hematopoietic progenitor cells in runners. *J Appl Physiol*. 2002;93:1691–7.
14. Clarke WL, Cox DJ, Gonder-Frederick LA, Julian D, Schlundt D, Polonsky W. Reduced awareness of hypoglycemia in adults with IDDM. A prospective study of hypoglycemic frequency and associated symptoms. *Diabetes Care*. 1995;18:517–22.
15. Campbell MD, Walker M, Trenell MI, Jakovljevic DG, Stevenson EJ, Bracken RM, et al. Large pre- and postexercise rapid-acting insulin reductions preserves glycemia and prevents early- but not late-onset hypoglycemia in patients with Type 1 diabetes. *Diabetes Care*. 2013;36:2217–24.
16. Campbell MD, Walker M, Trenell MI, Luzio S, Dunseath G, Tuner D, et al. Metabolic implications when employing heavy pre- and post-exercise rapid-acting insulin reductions to prevent hypoglycaemia in type 1 diabetes patients: a randomised clinical trial. *PLoS One*. 2014;9, e97143.
17. Campbell MD, Walker M, Trenell MI, Stevenson EJ, Turner D, Bracken RM, et al. A low-glycemic index meal and bedtime snack prevents postprandial hyperglycemia and associated rises in inflammatory markers, providing protection from early but not late nocturnal hypoglycemia following evening exercise in type 1 diabetes. *Diabetes Care*. 2014;37:1845–53.
18. West DJ, Morton RD, Bain SC, Stephens JW, Bracken RM. Blood glucose responses to reductions in pre-exercise rapid-acting insulin for 24 h after running in individuals with type 1 diabetes. *J Sports Sci*. 2010;28:781–8.
19. Rabasa-Lhoret R, Bourque J, Ducros F, Chiasson JL. Guidelines for premeal insulin dose reduction for postprandial exercise of different intensities and durations in type 1 diabetic subjects treated intensively with a basal-bolus insulin regimen (ultralente-lispro). *Diabetes Care*. 2001;24:625–30.

20. Medicine ACoS. ACSM's guidelines for exercise testing and prescription. Philadelphia: Lippincott Williams & Wilkins; 2013.
21. Van Craenenbroeck EM, Van Craenenbroeck AH, van Ierssel S, Bruyndonckx L, Hoymans VY, Vrints CJ, et al. Quantification of circulating CD34+/KDR+/CD45dim endothelial progenitor cells: analytical considerations. *Int J Cardiol.* 2013;167:1688–95.
22. Fadini GP, Baesso I, Albiero M, Sartore S, Agostini C, Avogaro A. Technical notes on endothelial progenitor cells: ways to escape from the knowledge plateau. *Atherosclerosis.* 2008;197:496–503.
23. Alessio AM, Beltrame MP, Nascimento MC, Vicente CP, de Godoy JA, Silva JC, et al. Circulating progenitor and mature endothelial cells in deep vein thrombosis. *Int J Med Sci.* 2013;10:1746–54.
24. Khan SS, Solomon MA, McCoy Jr JP. Detection of circulating endothelial cells and endothelial progenitor cells by flow cytometry. *Cytometry B Clin Cytom.* 2005;64:1–8.
25. Fadini GP, Albiero M, Vigili de Kreutzenberg S, Boscaro E, Cappellari R, Marescotti M, et al. Diabetes impairs stem cell and proangiogenic cell mobilization in humans. *Diabetes Care.* 2013;36:943–9.
26. Yue WS, Lau KK, Siu CW, Wang M, Yan GH, Yiu KH, et al. Impact of glycemic control on circulating endothelial progenitor cells and arterial stiffness in patients with type 2 diabetes mellitus. *Cardiovasc Diabetol.* 2011;10:113.
27. Antonio N, Fernandes R, Soares A, Soares F, Lopes A, Carvalheiro T, et al. Reduced levels of circulating endothelial progenitor cells in acute myocardial infarction patients with diabetes or pre-diabetes: accompanying the glycemic continuum. *Cardiovasc Diabetol.* 2014;13:101.
28. Hortenhuber T, Rami-Mehar B, Satler M, Nagl K, Hobaus C, Hollerl F, et al. Endothelial progenitor cells are related to glycemic control in children with type 1 diabetes over time. *Diabetes Care.* 2013;36:1647–53.
29. Spinetti G, Cordella D, Fortunato O, Sangalli E, Losa S, Gotti A, et al. Global remodeling of the vascular stem cell niche in bone marrow of diabetic patients: implication of the microRNA-155/FOXO3a signaling pathway. *Circ Res.* 2013;112:510–22.
30. Cesari F, Maruccci R, Gori AM, Burgisser C, Francini S, Sofi F, et al. Impact of a cardiac rehabilitation program and inflammatory state on endothelial progenitor cells in acute coronary syndrome patients. *Int J Cardiol.* 2013;167:1854–9.
31. Agnoletti L, Curello S, Bachetti T, Malacarne F, Gaia G, Comini L, et al. Serum from patients with severe heart failure downregulates eNOS and is proapoptotic: role of tumor necrosis factor-alpha. *Circulation.* 1999;100:1983–91.
32. Spinelli FR, Metere A, Barbati C, Pierdominici M, Iannuccelli C, Lucchino B, et al. Effect of therapeutic inhibition of TNF on circulating endothelial progenitor cells in patients with rheumatoid arthritis. *Mediat Inflamm.* 2013;2013:537539.
33. Chang E, Paterno J, Duscher D, Maan ZN, Chen JS, Januszzyk M, et al. Exercise induces stromal cell-derived factor-1alpha-mediated release of endothelial progenitor cells with increased vasculogenic function. *Plast Reconstr Surg.* 2015;135:340e–50e.
34. Francescato MP, Stel G, Geat M, Cauci S. Oxidative stress in patients with type 1 diabetes mellitus: is it affected by a single bout of prolonged exercise? *PLoS One.* 2014;9, e99062.
35. Oikonomou D, Kopf S, von Bauer R, Djuric Z, Cebola R, Sander A, et al. Influence of insulin and glargine on outgrowth and number of circulating endothelial progenitor cells in type 2 diabetes patients: a partially double-blind, randomized, three-arm unicenter study. *Cardiovasc Diabetol.* 2014;13:137.

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